

## ANTIBACTERIAL ACTIVITY OF *RHUS CORIARIA*. L EXTRACTS GROWING IN PALESTINE

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**Abstract:** Disk diffusion and micro-dilution techniques were used to determine the antibacterial activity of water, ethanolic and methanolic extracts of *Rhus coriaria* ripe berries against five clinical bacterial strains (Methicillin-resistant *Staphylococcus aureus* (MRSA), multi-drug resistant *Pseudomonas aeruginosa*, enterohemorrhagic *Escherichia coli* O157 (EHEC), *Proteus vulgaris* and *Klebsiella pneumoniae*). *Bacillus subtilis* ATCC6633 was used as a reference strain. The zone of inhibition varies depending on bacterial species and type of extract. The average diameter of inhibition zones ranges from 0 to 19 mm, 15 to 25 mm and 15 to 22 mm for water, ethanolic and methanolic extract, respectively. The MIC value of ethanolic extract was 1.25 mg/ml against MRSA, EHEC, *P. aeruginosa* and *Proteus vulgaris*, while 0.156 mg/ml against *B. subtilis*. In general these results showed that the antibacterial activity of *R. coriaria* ripe berries extracts was more effective against Gram-positive bacteria than Gram-negative.

**Running title:** antibacterial activity of *Rhus coriaria*

**Keywords:** *Rhus coriaria*, sumac, antibacterial activity, Palestine

## ANTIBACTERIAL ACTIVITY OF...

### Introduction

Plants are able to produce different compounds that be used to protect themselves against different types of pathogens <sup>(1)</sup>. Interest in medicinal plants has revived as a consequence of current problems associated with the use of antibiotics <sup>(2)</sup>. Spices are mainly used in foods mainly because they give desirable flavors and aromas; in addition it shows antimicrobial activity <sup>(3)</sup>.

Medicinal plants are important elements of indigenous medical systems in Palestine as well as in other developing countries. *R. coriaria* (Anacardiaceae) commonly known as sumac (also spelled as sumach), is a wild bush that grows in all Mediterranean areas, including Palestine. Phytochemicals in *R. coriaria* are being used as antibacterial, antidiarrheic, antidysenteric, antihepatotoxic, antiseptic, antispasmodic, antiviral, astringent, candidicide, hepatoprotective, hepatotonic, protisticide, analgesic, antigastric, anti-inflammatory, antioxidant, antiulcer, fungicide, cyclooxygenase-inhibitor and lipoxygenase inhibitor due to their contents of ellagic acid, gallic acid, isoquercitrin, myricitrin, myricetin, quercetin, quercitrin and tannic acid <sup>(4)</sup>.

The acidic tasty of *R. coriaria* fruits is made into a condiment and sour drink in the Middle East dishes. Its sour taste is derived form the citric and malic acids found in its juice. In Palestine, *R. coriaria* is a well-known spice, popular and has been utilized extensively in many different meals, such as in zater (dukka) which is a blend of sumac, thyme and citric acid with sesame seeds; almusakhan which is composed from fragmented chicken, small fragments of onions and sumac, as well as in salads and others.

Due to lack information about the effect of *R. coriaria* seeds on bacterial activity <sup>(5-7)</sup> this study was conducted in order to study the antimicrobial effect of water, methanolic and ethanolic extracts of *R. coriaria* against five clinical species of bacteria.

### Materials and Methods:

**Extract preparation:** *R. coriaria* fruits were collected from plants grown in mountains in North of Palestine. The taxonomic identity of the plants was confirmed by us as described by Flora Palaestina <sup>(8)</sup>. Air-dried and powdered of *R. coriaria* ripe berries (30 g) were extracted with water, 80% methanol and 80% ethanol; the extracts were filtered through Watman No. 2 filter paper under suction. Extracts were concentrated to dryness in vacuum and weighed.

**Test Bacteria:** Five bacterial strains isolated from clinical material from patients were used in the study: Methicillin-resistant *Staphylococcus aureus*

(MRSA), multi-drug resistant *Pseudomonas aeruginosa*, enterohemorrhagic *Escherichia coli* O157 (EHEC), *Proteus vulgaris* and *Klebsiella pneumoniae*. A reference strain [*Bacillus subtilis* ATCC6633] was also tested.

**Antimicrobial activity:** The antimicrobial activity was determined by the well diffusion method<sup>(9)</sup>. Wells of (6 mm diameter) were made in Mueller Hinton agar. Plates were seeded with a 24 h old culture of the bacterial strains. Plant extracts were added to the wells, a concentration of 5 mg/well. Triplicates of each concentration for each bacteria species were prepared. The inoculated plates were incubated at 37 °C for 24 h. The diameter of the inhibition zones were measured for each plate and the average reading of the three replicates for each antibacterial species are shown in Table 1. The standard tetracycline disk (30 µg) was used as a control.

Minimum inhibitory concentration (MIC) was determined by the micro-dilution method<sup>(10)</sup>. The reconstituted ethanol extract was diluted in Mueller Hinton broth to give a final concentration of 0.0195, 0.0391, 0.078, 0.156, 0.313, 0.625, 1.25, 2.5, 5, 10 mg/ml. The bacterial inoculum size was adjusted to the turbidity of the 0.5 #1McFarland standard so as to deliver a final inoculum of approximately 10<sup>5</sup> colony-forming units (CFU/ml). Using a micropipette, 50µl of the standard microbial broth culture were introduced into the wells. The test plates were incubated at 37 °C for 18 h. The endpoint (MIC) is taken as the lowest concentration of drug at which the microorganism tested does not show visible growth.

#### **Results:**

The antibacterial activity of *R. coriaria* seed extracts are shown in Table 1. The inhibition zones vary depending on bacterial species and type of extract. The average diameter of inhibition zones ranges from 0 to 19 mm, 15 to 25 mm and 15 to 22 mm for *R. coriaria* seeds water, ethanolic and methanolic extract, respectively. The largest diameter of inhibition zone was observed from ethanolic and methanolic extracts on the growth of MRSA and *B. subtilis*. No effects were detected for water extract on *P. vulgaris*, *P. aeruginosa* and *K. pneumoniae*. The MIC of ethanolic extract was 1.25 mg/ml against MRSA, EHEC, *P. aeruginosa* and *Proteus vulgaris*, while 0.156 mg/ml against *B. subtilis*.

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**Table 1. The antibacterial activity of water, ethanol and methanol extracts of the dried ripe berries of *Rhus coriaria* species**

Name of organisms	Radius of Inhibition in mm (mean of three tests)			Antibiotic Tetracycline (30 µg)
	Water	Ethanol	Methanol	
MRSA	19	25	22	30
<i>Bacillus subtilis</i>	16	23	22	14
EHEC	10	16	16	38
<i>Proteus vulgaris</i>	0	16	15	27
<i>P. aeruginosa</i>	0	15	15	12
<i>K. pneumoniae</i>	0	15	15	30

Abbreviations: MRSA -Methicillin-resistant *Staphylococcus aureus*; EHEC-Enterohemorrhagic *Escherichia coli* O157.

### Discussion

Traditional medicine is an important source of products for developing countries in treating common infectious bacteria. The emergence of multi-drug resistant infectious bacteria, high cost of synthetic compounds as well as undesirable side effects of certain drugs insist on pharmaceutical companies to look for new therapeutic agents from other alternative sources including medicinal plants. In recent years, different reports, from different countries were published showing the antimicrobial activities of medicinal plants <sup>(6, 11-24)</sup>. The result of the present study indicates the existence of antimicrobial activity in the crude extracts of *R. coriaria* seeds. Water extract was more active against gram-positive than gram-negative bacteria. Similar observations were reported elsewhere <sup>(7, 25)</sup>.

Ethanol and methanol extract displayed broad spectrum of activity, since Gram positive bacteria including Methicillin-resistant *S. aureus* and Gram negative bacteria including *P. aeruginosa*, enterohemorrhagic *E. coli* O157, *P. vulgaris* and *K. pneumoniae* were inhibited with the *R. coriaria* seed extracts. These results in agreement with those previously published <sup>(6, 11, 20)</sup>. *B. subtilis* was clearly found to be the most sensitive among them demonstrating a MIC 0.156 mg/ml. The rest of the species found to be more resistant with MIC of 1.25 mg/ml. Increasing the concentration of extracts will lead to increase inactivation of cells in all strains tested <sup>(8-7)</sup>. The antimicrobial activity demonstrated by sumac extract against the microorganisms tested in this study is comparable with those of other spices reported in the literature which inhibited the growth of Gram-positive organisms <sup>(20, 26-28)</sup>. Traditional using of this spice may help in protecting from several bacterial diseases spontaneously and may aid in control of bacterial growth in foods.

## References

1. Cowam MM., 1999 - *Plant Products as Antimicrobial Agents*. Clinical Microbiology Reviews, 12: 564–582.
2. Emori TG., Gaynes RP., 1993 - *An overview of nosocomial infections, including the role of the microbiology laboratory*. Clinical Microbiology Reviews, 6: 428–442.
3. Beuchat LR., Golden DA., 1989 - *Antimicrobials occurring naturally in foods*. Food Technology, 43: 134–142.
4. Duke JA., JoBogenschutz-Godwin M., DuCellier J., Duke P-A K., **CRC Handbook of Medical Plant**. CRC press, Boca Raton, 2003, 269-270.
5. Iauk L., Caccamo F., Speciale AM., Tempera G., Ragusa S., Panté G., 1998 - *Antimicrobial activity of Rhus coriaria L. leaf extract*. Phytotherapy Research, 12: S152-S153.
6. Nimri LF., Meqdam MM., Alkofahi A., 1999 - *Antibacterial Activity of Jordanian Medicinal Plants*. Pharmaceutical Biology, 37: 196 -201.
7. Nasar-Abbas SM., Kadir Halkman AK., 2004 - *Antimicrobial effect of water extract of sumac (Rhus coriaria L.) on the growth of some food borne bacteria including pathogens*. International Journal of Food Microbiology, 97: 63-69.
8. Zohary M., **Flora Palaestina**. Goldberg's Press, Jarusalem, 1972, part two. p 300.
9. National Committee for Clinical Laboratory Standards (NCCLS), **Performance standards for antimicrobial disk susceptibility tests**, NCCLS, Pennsylvania, USA, 1993, M2-A5.
10. National Committee for Clinical Laboratory Standards (NCCLS), **Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically**, NCCLS, Pennsylvania, USA, 2000, N7-A5.
11. Grosvenor PW., Supriono A., Gray DO., 1995 - *Medicinal plants from Riau Province, Sumatra, Indonesia. Part 2: Antibacterial and antifungal activity*. Journal of Ethnopharmacology, 45: 97–111.
12. Ratnakar P., Murthy PS., 1995 - *Purification and mechanisms of action of antitubercular principle from garlic (Allium sativum) active against isoniazid susceptible and resistant Mycobacterium tuberculosis H37RV*. Indian Journal of Clinical Biochemistry, 10: 14–18.
13. Silva O., Duarte A., Cabrita J., Gomes E., 1996 - *Antimicrobial activity of Guinea -Bissau traditional remedies*. Journal of Ethnopharmacology, 50: 55–59.
14. David M., 1997 - *Antimicrobial activity of garlic*. Antimicrobial Agents and Chemotherapy, 41: 2286.

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15. Saxena K., 1997 - *Antimicrobial Screening of Selected Medicinal Plants from India*. Journal of Ethnopharmacology, 58: 75–83.
16. Saxena VK., Sharma RN., 1999 - *Antimicrobial activity of essential oil of Lantana aculeata*. Fitoterapia, 70: 59–60.
17. Rojas G., Le´varo J., Tortoriello J., Navarro V., 2001 - *Antimicrobial evaluation of certain plants used in Mexican traditional medicine for the treatment of respiratory diseases*. Journal of Ethnopharmacology, 74: 97–101.
18. Rojas R., Bustamante B., Bauer B., Fern´ande I., Alb´an J., Lock, O., 2003 - *Antimicrobial activity of selected Peruvian medicinal plants*. Journal of Ethnopharmacology, 88: 199–204.
19. Ahmad I., Beg AZ., 2001 - *Antimicrobial and phytochemical studies on 45 Indian medicinal plants against multi-drug resistant human pathogens*. Journal of Ethnopharmacology 74: 113–123.
20. Srinivasan D., Nathan S., Suresh T., Perumalsamy PL., 2001 - *Antimicrobial activity of certain Indian medicinal plants used in folkloric medicine*. Journal of Ethnopharmacology 74: 217–220.
21. Tshibangu JN., Chifundera K., Kaminsky R., Wright AD., Gabriele Maria Ko`nig GM., 2002 - *Screening of African medicinal plants for antimicrobial and enzyme inhibitory activity*. Journal of Ethnopharmacology 80: 25–35.
22. Machado TB., Pinto AV., Pinto MCFR., Leal ICR., Silva MG., Amaral ACF., Kuster RM., Netto-dosSantos KR., 2003 - *In vitro activity of Brazilian medicinal plants, naturally occurring naphthoquinones and their analogues, against methicillin-resistant Staphylococcus aureus*. International Journal of Antimicrobial Agents, 21: 279-284.
23. Karaman. I., Sahin F., Gllce M., Ogtu H., Sengl M., Adgzel A., 2003 - *Antimicrobial activity of aqueous and methanol extracts of Juniperus oxycedrus L*. Journal of Ethnopharmacology, 85: 231–235.
24. Voravuthikunchai S., Lortheeranuwat A., Jeeju W., Sririrak T., Phongpaichit S., Supawita T., 2004 - *Effective medicinal plants against enterohaemorrhagic Escherichia coli O157:H7*. Journal of Ethnopharmacology, 94: 49–54.
25. Marino M., Bersani C., Comi M., 1999 - *Antimicrobial activity of the essential oils of Thymus vulgaris L. measure using a bioimpedimetric method*. Journal of Food Protection, 62: 1017– 1023.
26. Shelef L.A., Naglik OA., Bogen BW., 1980 – *Sensitivity of some common food borne bacteria to the spices sage, rosemary and all spice*. Journal of Food Science, 45: 1042–1044.

27. Ting WTE., Deibel KE., 1992 - *Sensitivity of Listeria monocytogenes to spices at two temperatures*. Journal Food Safety, 12: 129– 137.
28. Jain R.C., 1994 - *Antibacterial activity of garlic extract*. Indian Journal of Medical Microbiology, 11: 26–31.