

ASSESSMENT OF PHENOTYPIC DIVERSITY OF BARLEY GENOTYPES THROUGH CLUSTER AND PRINCIPAL COMPONENT ANALYSES

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ABSTRACT

Determination of genetic diversity is useful for plant breeding and hence production of more efficient plant varieties under different conditions. Accordingly, a collection of 74 accessions of landraces and cultivated varieties of barley from different countries, mainly from the Fertile Crescent were selected, grown and analyzed for phenotypic diversity. The field experiment was conducted at the experimental farm of the Faculty of Agriculture, An-Najah National University, Tulkarm (Khadouri), Palestine in a randomized complete block design with three replications. Initially, an analysis of variance (ANOVA) was conducted to test for significant differences among barley accessions in measured traits. A two-step cluster analysis was performed using the eleven measured traits to determine the optimal number of clusters based on Schwarz's Bayesian Criterion (BIC) then, a dendrogram was constructed using the Hierarchical Cluster analysis with Ward's clustering method based on Squared Euclidean Distances. ANOVA revealed highly significant differences among barley accessions in all studied traits. Based on Principal Component Analysis (PCA), the first four extracted components explained 76.1% of the total variation in the 11 studied traits. The clustering analyses revealed two main clusters each can be further divided into two sub-clusters. The first cluster included 41 accessions and the second cluster included 33 accessions. Such variation among studied accessions can be utilized in designing new breeding programs and crossing nurseries for barley improvement.

Key words: Cluster analysis, *Hordeum vulgare*, PCA, Selection, Barley

Published first online January 21, 2021.

Published final August 07, 2021.

INTRODUCTION

Barley (*Hordeum vulgare* L.) is one of the most important cereals currently cultivated in the world. It is considered as one of the main important sources of protein and calories in human diet. Historically, barley is one of the oldest domesticated grains in the world. Its cultivation started between 9500 and 8400 years ago and it played a vital role in the revolution of civilizations by providing food to humans and animals (Azhaguvel and Komatsuda, 2007). Barley, wheat and several pulses (grain legumes) were originated in the 'Fertile Crescent', specifically Palestine -Jordan area. This area is the region in which barley was brought into culture and then spread through Syria and Lebanon to northern Iraq and Iran (Preece *et al.*, 2016).

Breeding for high yielding varieties generally leads to reduce genetic diversity that can change gene frequencies of plant material (Malik *et al.*, 2013). Knowledge regarding the amount of variation in germplasm arrays and relationships between genotypes are important considerations for efficient conservation and utilization of genetic resources (Russel *et al.*, 1997; Davila *et al.*, 1998 and Manjunatha *et al.*, 2006). In the

context of plant improvement, this information provides a basis for making decisions regarding selection of parental combinations that amount of genetic variation present, and the location of the genetic determinants of diversity may be useful for germplasm conservation and targeting gene discovery efforts (Sorrels and Wilson, 1997; Jana, 1999 and Hou *et al.*, 2005). It is, therefore, important to study variability in plant genotypes to meet the diversified goals such as increasing yield, wider adaptation, desirable quality, and pests and disease resistance (Fufa *et al.*, 2005). Growing numbers of candidate varieties and the decrease in variability in morphological traits has led to the establishment of evaluation procedures to discriminate accessions during germplasm evaluation (Aghaee *et al.*, 2010).

Multivariate analysis is the most commonly used approach to illuminate the patterns of variation in germplasm collections. Among multivariate techniques, PCA and cluster analysis are preferred tools for morphological characterization of genotypes and their grouping on similarity basis (Mohammadi and Prasanna, 2003 and Peeters and Martinelli, 1989). Combination of these two approaches give comprehensive information of characters which are critically contributing for genetic

variability in crops (Rachovska *et al.*, 2003). The present study was undertaken with the objective to assess and evaluate the diversity of 74 accessions of barley based on agro-morphological traits.

MATERIALS AND METHODS

Plant Material: A collection of 74 accessions of landraces and cultivated varieties of barley from different countries, mainly from the Fertile Crescent, kindly provided by Dr. Maria von Korff, Max-Planck Institute for Plant Breeding, Germany, was used in the experiment (Table 1).

Field Experiment: The field experiment was conducted at the experimental farm of the Faculty of Agriculture, An-Najah National University, Tulkarm (Khadouri),

Palestine (32.31519° N and 35.02033° W and altitude of 75 m, average mean yearly rainfall 600 mm), during two growing seasons 2015-2016 and 2016-2017 in a triplicated randomized complete block design (RCBD). In each replicate, twenty seeds from each accession were planted in one-meter row. Spacing was 10 cm between plants within row and 70 cm between rows.

Data collection: Observations were recorded on five plants from each replicate on each accession. The traits measured were growth vigor (measured on a scale from 1 = very low to 5 = very high), days to stem elongation, days to heading, days to maturity, number of tillers per plant, spike length (cm), spike number, plant height (cm), vegetative biomass (g), thousand-kernel weight (g) and grain yield per row (g).

Table 1: Barley accessions used in the study.

NO	Code/name	NO	Code/name	NO	Code/name	NO	Code/name
1	MK_RB_18	20	MK_RB_183	39	MK_RB_246	58	LR1897
2	MK_RB_21	21	MK_RB_184	40	MK_RB_268	59	Barke
3	MK_RB_86	22	MK_RB_186	41	MK_RB_269	60	Lr761
4	MK_RB_87	23	MK_RB_187	42	MK_RB_270	61	Optic
5	MK_RB_94	24	MK_RB_188	43	MK_RB_271	62	HID44
6	MK_RB_107	25	MK_RB_189	44	MK_RB_278	63	HID52
7	MK_RB_113	26	MK_RB_190	45	MK_RB_279	64	HID301
8	MK_RB_114	27	MK_RB_192	46	MK_RB_281	65	LR1043
9	MK_RB_118	28	MK_RB_223	47	MK_RB_282	66	Marthe
10	MK_RB_147	29	MK_RB_224	48	MK_RB_284	67	Bowman
11	MK_RB_150	30	MK_RB_225	49	MK_RB_286	68	BW281
12	MK_RB_152	31	MK_RB_227	50	Mutha	69	BW284
13	MK_RB_154	32	MK_RB_228	51	Rum	70	BW285
14	MK_RB_155	33	MK_RB_229	52	Aksad	71	BW287
15	MK_RB_156	34	MK_RB_230	53	Keel	72	BW289
16	MK_RB_157	35	MK_RB_232	54	Flagship	73	BW290
17	MK_RB_163	36	MK_RB_233	55	Morex	74	G400
18	MK_RB_167	37	MK_RB_240	56	Auriga		
19	MK_RB_181	38	MK_RB_241	57	LR871		

Data Analysis

Analysis of Variance (ANOVA): Initially, an analysis of variance (ANOVA) (Fisher, 1918) was conducted using PROC GLM procedure of SAS/STAT software, version 9.0 for Windows (SAS institute 2002) to test differences among barley accessions in measured traits. The analysis model included the effects of year, replicate, and accession. For each trait, the observed means (averages over all replicates and over the two growing seasons) were obtained for each genotype and used in the subsequent analyses.

Principal Component Analysis: Factor analysis with Principal Components (Pearson, 1901 and Hotelling, 1933) was carried out in SPSS (V21.0). KMO (Kaise-

Meyer-Olkin Measure of Adequacy) test value of 0.59 and the significant result of Bartlett's test of Sphericity ($P < 0.001$) indicated that PCA multivariate analysis is appropriate for the data. Rotated solutions of principal components were obtained using Oblimin with Kaiser Normalization method (Kaiser, 1958; Jennrich and Sampson, 1966; and Clarkson and Jennrich, 1988)

Cluster Analysis: First, a two-step cluster analysis (Chiu *et al.*, 2001 and Bacher *et al.*, 2004) was performed on the barley accessions using the eleven measured traits. This initial analysis was done to determine the optimal number of clusters based on Schwarz's Bayesian Criterion (BIC) and determine the relative importance of the measured traits in clustering of the studied accessions.

Then, a Hierarchical Cluster analysis with Ward's clustering method (Ward, 1963) based on Squared Euclidean Distances was performed to construct a cluster tree (dendrogram). Student's t test (Gosset, 1908) was applied to test for differences in means of measured traits between the two main clusters which were revealed by the clustering analyses. Clustering analyses and the t test were all carried out in SPSS (V21.0).

RESULTS AND DISCUSSION

Analysis of Variance: The results from the analysis of variance are in Table 2. The effect of year was highly significant ($P < 0.0001$) for all traits (except for growth vigor) reflecting high environmental variation between the two growing seasons. The effect of block was not significant except for growth vigor ($P < 0.0001$), plant height ($P = 0.05$), spike number ($P = 0.004$) and thousand-kernel weight ($P < 0.0001$). The results showed highly significant differences ($P < 0.0001$) among accessions for all studied traits. This large variation among genotypes could be utilized in selection programs particularly for production traits.

Principal Component Analysis: Although eleven principal components could have been extracted (equal to number of traits), only the first four components were considered important (had Eigenvalues above 1.0). These

results are in agreement with the results reported by Maqbool *et al.* (2010). These four components explained 76.1% of the total variation in the 11 studied traits (Table 3 and Figure 1). The components plot (Figure 2) and the patterns matrix (Table 4) showed the contribution of studied traits to extracted components. Characters with absolute values closer to unity have higher contribution to the components (Chahal and Gosal, 2002).

The first component which explained 27% of the total variation was dominated by three traits with high positive loadings (days to stem elongation, days to heading and days to maturity) and by growth vigor which has a negative contribution. The second component which explained 20% of the total variation was dominated by three traits (tiller number and spike number with positive loadings and plant height with a negative loading). The third component explained 18% of the total variation and had high positive loadings for plant height, grain yield and vegetative biomass. The fourth component explained about 11% of the total variation and had high positive loadings for spike length and thousand-kernel weight. These were the major effective traits that governed the variation in these four components. Chahal and Gosal (2002) and Poudel *et al.*, (2017), stated that characters with largest absolute values closer to unity within the first PC influence the clustering more than those with lower absolute values closer to zero.

Table 2. Analysis of variance results (mean squares) of data on seventy-four barley accessions. The model included the effects of year, block and accession.

Trait	Effects fitted in the model					
	Year		Block		Accession	
	Mean square	P value	Mean square	P value	Mean square	P value
Growth vigor	0.036	0.85	6.29	0.002	3.11	< 0.0001
Days to stem elongation	4314.1	< 0.0001	176.3	0.30	1030.6	< 0.0001
Days to heading	20229.8	< 0.0001	18.0	0.47	1352.9	< 0.0001
Days to maturity	31621.6	< 0.0001	16.3	0.44	677.5	< 0.0001
Tiller number	5874.4	< 0.0001	29.7	0.28	99.0	< 0.0001
Spike number	3920.8	< 0.0001	137.49	0.004	92.3	< 0.0001
Spike length	37.9	< 0.0001	1.3	0.10	4.4	< 0.0001
Plant height	12423.5	< 0.0001	284.1	0.05	721.1	< 0.0001
Grain yield	262391.4	< 0.0001	2757.67	0.35	22637.8	< 0.0001
Thousand-kernel weight	371.1	< 0.0001	128.8	< 0.0001	202.3	< 0.0001
Vegetative biomass	37723015.1	< 0.0001	232.8	0.99	297292.2	< 0.0001

Cluster Analysis: The clustering analyses revealed two main clusters each can be further divided into two sub-clusters (Figure 3). The first cluster included 41 accessions and the second cluster included 33 accessions. Similar results were reported in a collection of 133 barley accessions from Pakistan (Zaheer *et al.*, 2008). The two-step cluster analysis showed that days to maturity, days to heading, and days to elongation were the most important traits in clustering the barley accessions (Figure 4)

confirming the results from the PCA analysis. Vegetative biomass, grain yield, and growth vigor had moderate importance in clustering the genotypes while the remaining traits (thousand-kernel weight, plant height, tiller number and spike number) were the least important in clustering the studied barley accessions. However, previous research showed that cluster analysis based on PCA is a more precise indicator of differences among wheat genotypes than cluster analysis (not based on PCA)

(Khodadadi *et al.*, 2011). Accessions in Cluster 1 had significantly higher means of days to stem elongation, days to heading and days to maturity and significantly lower means of grain yield, growth vigor, spike number, tiller number, thousand-kernel weight, and vegetative

biomass (Table 5). Plant height and spike length did not differ between the two clusters. Similar works have been done by Maqbool *et al.* (2010), Degewione and Alamerew (2013) and Sajjad *et al.* (2011) for grouping of wheat germplasm by principal component analysis.

Table 3. Eigenvalues and percentage of total variance explained by each principal component.

Component	Initial Eigenvalues			Extraction Sums of Squared Loadings		
	Total	% of Variance	Cumulative %	Total	% of Variance	Cumulative %
1	2.973	27.024	27.024	2.973	27.024	27.024
2	2.222	20.200	47.224	2.222	20.200	47.224
3	1.980	18.003	65.227	1.980	18.003	65.227
4	1.195	10.862	76.089	1.195	10.862	76.089
5	.747	6.790	82.878			
6	.736	6.688	89.566			
7	.537	4.879	94.445			
8	.323	2.937	97.382			
9	.188	1.706	99.088			
10	.063	.571	99.658			
11	.038	.342	100.000			

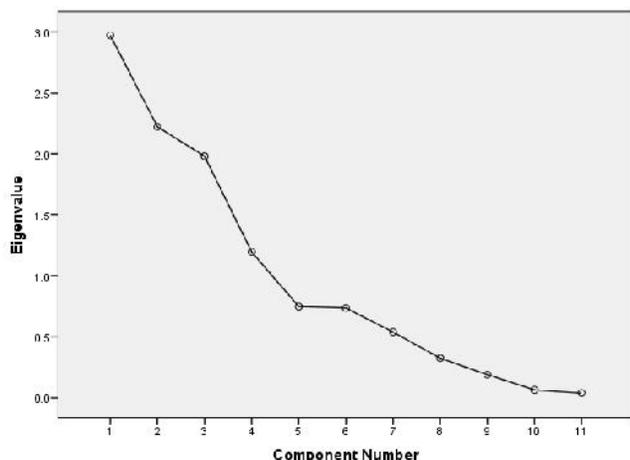


Figure 1. Scree plot of principal components and their Eigenvalues.

Table 4. The pattern matrix from Principal Component Analysis showing the contributions (loadings) of measured traits to the first four extracted components.

Trait	Principal Component			
	1	2	3	4
Growth vigor	-.521	-.048	.167	.287
Days to stem elongation	.822	.208	-.140	.006
Days to heading	.951	-.127	.110	.136
Days to maturity	.951	-.101	.133	.056
Tiller number	-.076	.937	.071	.122
Spike number	.086	.947	.210	.015
Spike length	.089	-.019	-.226	.812
Plant height	.049	-.584	.478	.083
Grain yield	.035	.097	.931	-.007
Thousand-kernel weight	-.052	.113	.155	.703
Vegetative biomass	-.064	.066	.896	-.069

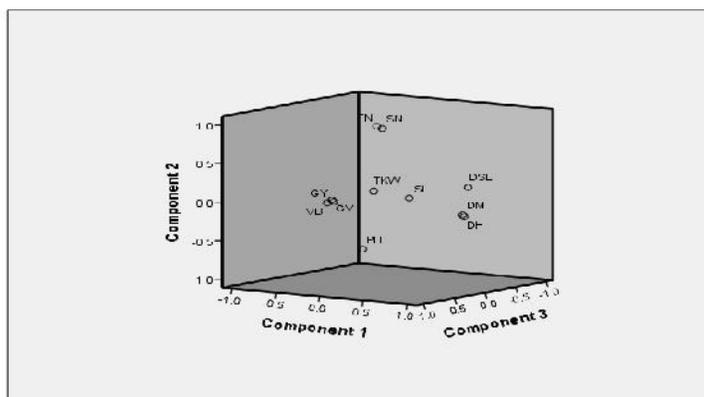


Figure 2. Component plot in rotated space showing the contribution of each of the eleven studied traits on barley genotypes (DH = days to heading, DM = days to maturity, DSE = days to stem elongation, GV = growth vigor, GY = grain yield, PH = plant height, SL= spike length, SN = spike number, TKW= thousand-kernel weight, TN= tiller number, VB = vegetative biomass).

Table 5. Means of studied traits by cluster.

Trait	Cluster		P value
	1	2	
Growth vigor	4.2	4.9	< 0.0001
Days to stem elongation	67.2	51.9	< 0.0001
Days to heading	103.7	83.9	< 0.0001
Days to maturity	123.2	109.7	< 0.0001
Tiller number	21.2	23.6	0.013
Spike number	17.9	19.8	0.033
Spike length, cm	8.1	7.8	0.27
Plant height, cm	71.2	75.6	0.085
Grain yield, g	125.2	185.5	< 0.0001
Thousand-kernel weight, g	39.8	43.3	< 0.0001
Vegetative biomass, g	473.6	684.6	< 0.0001

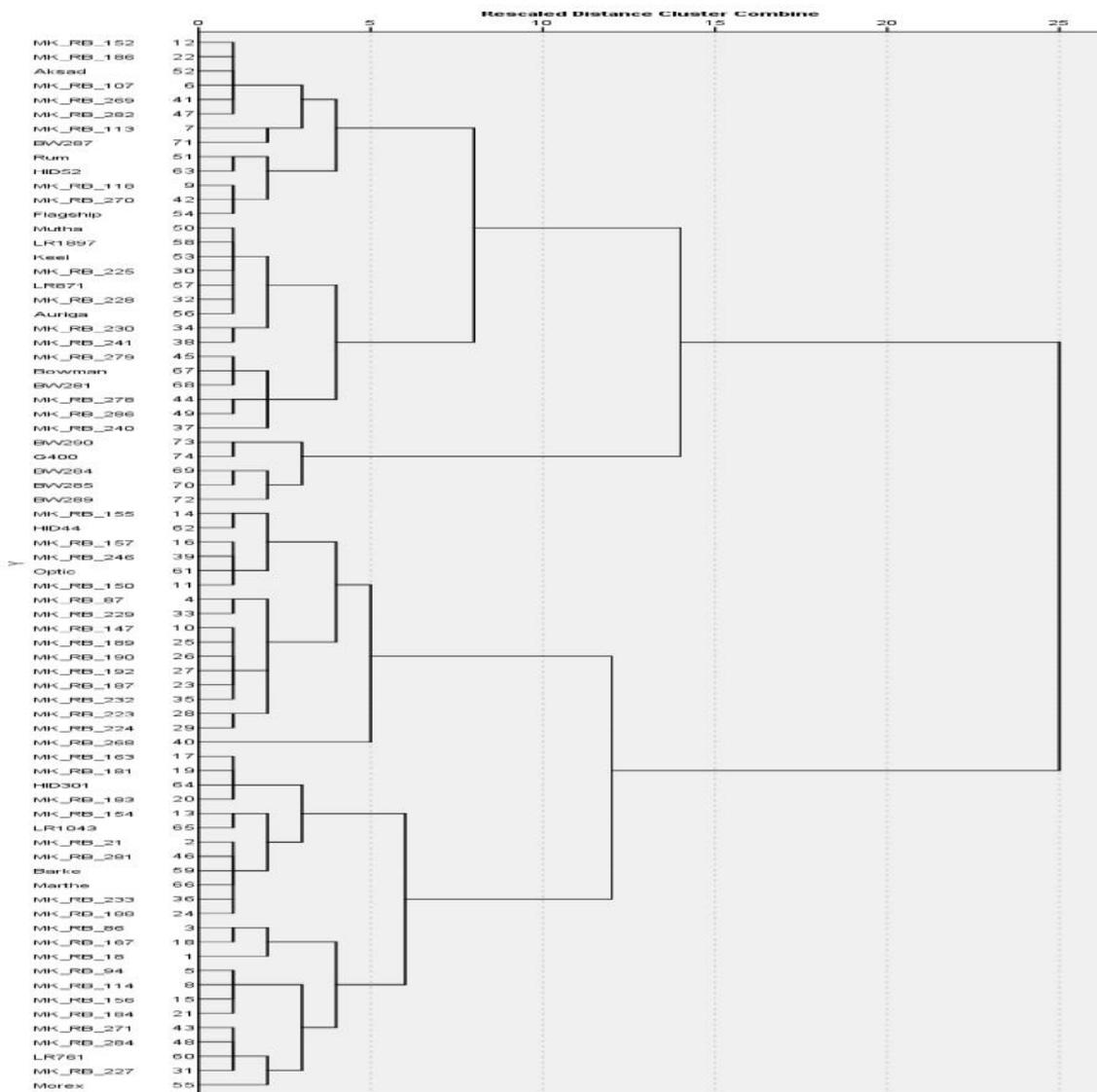


Figure 3. Dendrogram of 74 Barley accessions using the Hierarchical Ward's clustering method based on 11 measured traits.

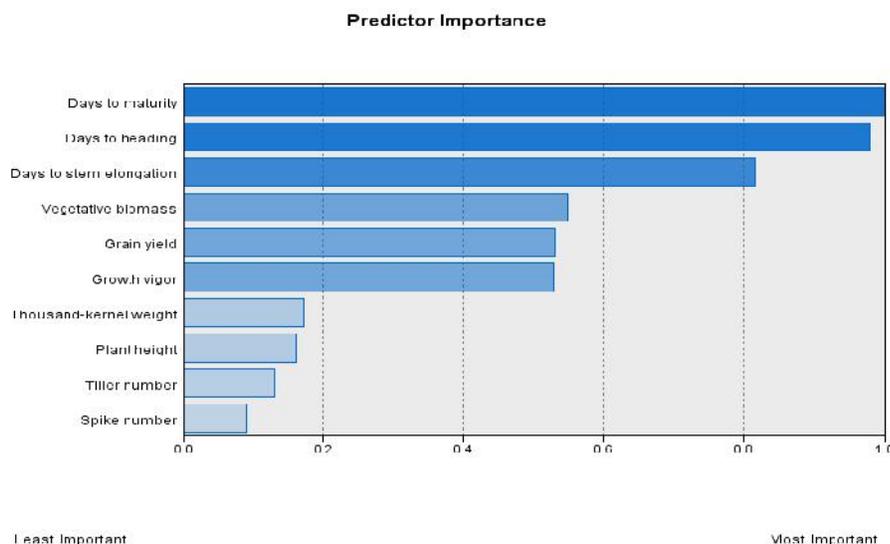


Figure 4. Relative importance of measured traits in clustering of studied genotypes.

Conclusions: The present study showed large amount of variation among studied genotypes for all measured characters indicating that high opportunities exist for genetic improvement of barley genotypes through direct selection and conservation of the germplasm for future utilization. These genotypes can be considered for breeding operations as well as for further study for developing superior barley genotypes. These barley genotypes need to be crossed and selected to develop high yielding pure line varieties.

Acknowledgments: This research was supported by An-Najah National University. The authors gratefully acknowledges Dr. Maria von Korff (Institute for Plant Genetics, Heinrich Heine University Düsseldorf, Germany) for providing seed samples and the excellent technical assistance by the technical team of the experimental farm of the Faculty of Agriculture, An-Najah National University

REFERENCES

- Aghaee, M., R. Mohammadi and S. Nabovati (2010). Agro-morphological characterization of durum wheat accessions using pattern analysis. *Aust. J. Crop Sci.* 4: 505-514.
- Azhaguvel, P. and T. Komatsuda (2007). Aphylogenetic analysis based on nucleotide sequence of a marker linked to the Brittle Rachis Locus indicates a diphyletic origin of barley. *Ann. Bot.* 100:1009–1015.
- Bacher, J., K. Wenzig and M. Vogler (2004). SPSS two step cluster - a first evaluation. *Univ. Erlangennürnberg*. 1, 1–20
- Chahal, G.S. and S.S. Gosal (2002). Principles and Procedures of Plant Breeding: Biotechnology and Conventional Approaches. Narosa Publishing House. New Delhi, India. pp. 604.
- Chiu, T., D. Fang, J. Chen, Y. Wang and C. Jeris (2001). A robust and scalable clustering algorithm for mixed type attributes in large database environment, in *Proceedings of The Seventh ACM SIGKDD International Conference on Knowledge Discovery and Data Mining - KDD '01*, (New York, NY: ACM Press), 263–268.
- Clarkson, D.B. and R.I. Jennrich (1988). Quartic rotation criteria and algorithms. *Psychometrika* 53, 251–259.
- Davila, J.A., M.P.S. Hoz, Y. Loarce and E. Ferrer (1998). The use of random amplified microsatellite polymorphic DNA and coefficients of parentage to determine genetic relationships in barley. *Genome* 41: 477-486.
- Degewione, A. and S. Alamerew. (2013). Genetic diversity in bread wheat (*Triticum aestivum* L.) genotypes. *Pakistan J. Biol. Sci.* 16: 1330-5
- Fisher, R. A. (1918). The correlation between relatives on the supposition of Mendelian inheritance. *Philosophical Transactions of the Royal Society of Edinburgh*, 52: 399–433.
- Fufa, H., P.S. Baenziger, B.S. Beecher, I. Dweikat, R.A. Graybosch and K. M. Eskridge (2005). Comparison of phenotypic and molecular marker-based classifications of hard red winter wheat varieties. *Euphytica* 145: 133-146.
- Gosset, W. S. (1908). The probable error of a mean. *Biometrika* 6(1): 1–25
- Hotelling, H. (1933). Analysis of a complex of statistical variables into principal components. *J. Educ. Psychol.* 24: 417-441.

- Hou, Y.C., Z.H. Yan, Y.M. Wei and Y.L. Zheng (2005). Genetic diversity in barley from West China. *Barley Genet. Newsl.* 35: 9-22.
- Jana, S. (1999). Some recent issues on the conservation of crop genetic resources in developing countries. *Genome* 44: 562-569.
- Jennrich, R.I. and P. F. Sampson (1966). Rotation for simple loadings. *Psychometrika* 31: 313-323.
- Kaiser, H.F. (1958). The varimax criterion for analytic rotation in factor analysis. *Psychometrika* 23: 187-200.
- Khodadadi, M., M.H. Fotokian and M. Miransari (2011). Genetic diversity of wheat (*Triticum aestivum* L.) genotypes based on cluster and principal component analyses for breeding strategies. *Aust. J. Crop. Sci.* 5: 17-24
- Malik. R., H. Sharma, A. Verma, S. Kundu, I. Sharma and R. Chatrath (2013). Hierarchical clustering of Indian wheat varieties using morphological diversity assessment. *Indian J. Agric. Res.* 47: 116-123.
- Manjunatha, T., I.S. Bisht, K.V. Bhat and B.P. Singh (2006). Genetic diversity in barley (*Hordeum vulgare* L.) landraces from Uttaranchal. *Genet. Resour. Crop. Evol.* 54: 55-65.
- Maqbool, R., M. Sajjad, I. Khaliq, Aziz-ur- Rehman, A.S. Khan, and S.H. Khan (2010). Morphological diversity and traits association in bread wheat (*Triticum aestivum* L.) *Am. Eurasian J. Agric. Environ. Sci.* 8: 216-224.
- Mohammadi, S.A. and B.M. Prasanna (2003). Analysis of genetic diversity in crop plants-salient statistical tools and considerations. *Crop Sci.* 43: 1235-1248.
- Pearson, K. (1901). On lines and planes of closest fit to systems of points in space, *Philosophical Magazine, Series 6*, 2, no. 11, pp. 559-572.
- Peeters, J.P., and J.A. Martinelli (1989). Hierarchical cluster analysis as a tool to manage variation in germplasm collections. *Theor. Appl. Genet.* 78: 42-48.
- Poudel, A., D.B. Thapa and M. Sapkota (2017). Assessment of genetic diversity of bread wheat (*Triticum aestivum* L.) genotypes through cluster and principal component analysis. *Inter. J. Exp. Res. Rev.* 11: 1-9
- Preece, C., A. Livarda, A. Christin, M. Wallace, G. Martin, M. Charles, G. Jones, M. Rees and P. Osborne (2016). How did the domestication of Fertile Crescent grain crops increase their yields? *Funct. Ecol.* 31: 387-397.
- Rachovska, G., D. Dimova and B. Bojinov (2003). Application of cluster analysis and principal component analysis for evaluation of common winter wheat genotypes. *Proceedings of the scientific session of jubilee 2002-Sadovo*, volume III: 68-72.
- Russel, J.R., J.D. Fuller, M. Macaulay, B.G. Hatz, A. Jahoor, W. Powell and R. Waugh (1997). Direct comparison of levels of genetic variation among barley accessions detected by RFLP, AFLPs, SSRs and RAPDs. *Theor. Appl. Genet.* 95: 714-722.
- Sajjad, M., S.H. Khan and S.S. Khan (2011). Exploitation of germplasm for grain yield improvement in spring wheat (*Triticum aestivum*). *Int. J. Agric Biol.* 13: 695-700.
- SAS Institute (2002). *User's Guide. Statistics. Ver.9.0.* Cary, N.C.
- Sorrels, M.E. and W.A. Wilson (1997). Direct classification and selection of superior alleles for crop improvement. *Crop Sci.* 37: 691-697.
- Ward, J. H. (1963). Hierarchical grouping to optimize an objective function. *J. Am. Stat. Assoc.* 58, 236-244.
- Zaheer, A., S.U. Ajmal, M. Munir, M. Zubair and M.S. Masood (2008). Genetic diversity for morpho-genetic traits in barley germplasm. *Pakistan J. Bot.* 40: 1217-1224.